



Photo by Juan González

Crosstalk between Ub family members under stress conditions

UBICODE & UBIRED TRAINING SESSION

MARCH 11-15, 2019

CENTRO DE INVESTIGACIÓN PRÍNCIPE FELIPE - VALENCIA, ES

GENERAL INFORMATION

Protein post-translational modification by ubiquitin (Ub) and Ubiquitin-like proteins (UBLs) is versatile, highly dynamic and involved in nearly all aspects of biological functions in eukaryotes. The reversibility and heterogeneity of Ub and UBL chains attached to protein substrates have complicated their isolation, quantification and characterization. Most approaches used to identify and characterise modified proteins are based in tagged versions of Ub and UBLs. Strategies have emerged to isolate endogenous ubiquitylated targets, including molecular traps based on the use of Ub-binding peptides, such as TUBEs (Tandem-repeated Ubiquitin-Binding Entities), peptide aptamers and affimers. Altogether these tools will contribute to unravel the secrets of the Ub-code in cell physiology and pathology. Knowing the benefits and limits of each of these approaches will give a better picture of the available technologies and help our fellows to choose the most appropriate tools to determine the modification of substrates with Ub/UBLs.

CONTENT

In this course we will use some of these tools and technologies to learn about their specificity, affinity and their applications. The capacity to capture and identify distinct chains with these tools will be analysed and compared. Strategies include His6 purification, pan or chain-selective molecular traps (to capture ubiquitin, SUMO and NEDD8) and chain specific nanobodies and/or minibodies.

TECHNOLOGY

Modified proteins will be captured using affinity chromatography. Captured material will be analysed by Western blot. Affinities of distinct tools for specific chains will be tested by thermophoresis (Nanotemper).

OBJECTIVES

To train students on techniques for the detection of total and individual proteins simultaneously modified by Ub and UBLs.

TRAINERS

Dimitris Xirodimas (CNRS-UbiCODE), Manuel S. Rodriguez (CNRS-UbiCODE), Laurie Bonnafé (CNRS), Rosa Farras (CIPF), Pierre Soule (Nanotemper), Frank Bringold (Nanotemper), Enrique Garcia Gomez (GE Healthcare)



PROGRAMME

MONDAY 11/03

Morning *Theory*

9:00

Welcome and tour de table

9:15

Objectives of the course

9:30

Introduction to Ub/UbLs signalling
(*Manuel Rodriguez*)

11:00

COFFEE BREAK offered by Werfen

11:30

Study of the response of Ub/UbL to proteotoxic stress (e.g. proteasome inhibitors) (*Dimitris Xirodimas*)

12:30

How to combine different chromatographic techniques in the purification of proteins
(*Enrique Garcia Gomez - GE Healthcare*)

13:30

LUNCH

Afternoon *Theory*

15:00

Study of the response of Ub/UbLs to proteotoxic stress (e.g. heat shock, proteasome inhibitors);
Methods to isolate and identify Ub/UbLs substrates;
Detection of hybrid Ub/UbL chains
(*Manuel Rodriguez, Dimitris Xirodimas*)

MONDAY 11/03

Late-afternoon
Practice

16:00

Cell lysis and capture of proteins modified with hybrid Ub/UbLs chains using His-6 tags and binding entities to capture Ub, SUMO and NEDD8

18:30

FREE EVENING

TUESDAY 12/03

Morning
Practice

9:00

Columns wash and samples elution

10:30

Gel electrophoresis

11:30

Gel transfer

13:25

Gel blocking

13:30

LUNCH

Afternoon
Practice

15:00

Preparation of antibodies
Overnight antibody incubation

Afternoon
Theory

16:00

Short presentation of students' individual projects (5 min.)

17:00

COFFEE BREAK

17:30

Analysis of the affinities of Ub or SUMO chains for binding peptides/proteins using various techniques

18:30

FREE EVENING

WEDNESDAY 13/03

Morning *Practice*

9:00	Membrane washing and 2nd antibody incubation
10:30	Development
12:00	Data collection and interpretation
13:30	LUNCH

Afternoon *Theory*

15:00	Microscale Thermophoresis (MST) part I (<i>Frank Bringold, Pierre Soule</i>)
16:00	COFFEE BREAK
16:30	MST part II
18:30	FREE EVENING

THURSDAY 14/03

Morning *Practice*

9:00	MST experiments including affinities of binding entities for Ubiquitin, SUMO, or nanobodies for K48 and K63 chains
13:00	LUNCH

Afternoon

15:00	MST data collection and analysis.
18:30	FREE TIME

20:30 - FAREWELL DINNER

FRIDAY 15/03

Morning	9:00	Data analysis and interpretation
	13:00	LUNCH
Afternoon	15:00	Final discussion

17:00 - SCHEDULED END OF THE COURSE

PRACTICAL INFORMATION



Venue - Centro de Investigación Príncipe Felipe (CIPF)

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